The Antituberculosis Drug Pyrazinamide Affects the Course of Cutaneous Leishmaniasis In Vivo and Increases Activation of Macrophages and Dendritic Cells

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Antileishmanial therapy is suboptimal due to toxicity, high cost, and development of resistance to available drugs. Pyrazinamide (PZA) is a constituent of short-course tuberculosis chemotherapy. We investigated the effect of PZA on Leishmania major promastigote and amastigote survival. Promastigotes were more sensitive to the drug than amastigotes, with concentrations at which 50% of parasites were inhibited (MIC50) of 16.1 and 8.2 μM, respectively (48 h posttreatment). Moreover, 90% of amastigotes were eliminated at 120 h posttreatment, indicating that longer treatments will result in parasite elimination. Most strikingly, PZA treatment of infected C57BL/6 mice resulted in protection against disease and in a 100-fold reduction in the parasite burden. PZA treatment of J774 cells and bone marrow-derived dendritic cells and macrophages increased interleukin 12, tumor necrosis factor alpha, and activation marker expression, as well as nitric oxide production, suggesting that PZA enhances effective immune responses against the parasite. PZA treatment also activates dendritic cells deficient in Toll-like receptor 2 and 4 expression to initiate a proinflammatory response, confirming that the immunostimulatory effect of PZA is directly caused by the drug and is independent of Toll-like receptor stimulation. These results not only are strongly indicative of the promise of PZA as an alternative antileishmanial chemotherapy but also suggest that PZA causes collateral immunostimulation, a phenomenon that has never been reported for this drug.

The leishmaniases are a group of insect-transmitted parasitic diseases prevalent worldwide, endemic in 88 countries; 350 million people are at risk, and 12 million people are affected. Two million new cases are estimated to occur annually, although only 600,000 are officially reported (10). During the last two decades, it has become increasingly apparent that the leishmaniases are much more prevalent than had been previously suspected. With human migration and vector expansion dramatically affecting the spread of disease, dramatic outbreaks have occurred in locations with previously low levels of infection (e.g., Kabul, Afghanistan, with more than 200,000 infected [22]). Immunologically naive individuals from the developed world traveling to areas of endemicity are particularly prone to infection. Leishmaniasis has been found among American soldiers deployed to the Middle East during both Gulf wars, current conflicts in Afghanistan, and Central America (3, 7, 12, 14, 18, 20, 24, 29). Civilians traveling into these areas are also at risk (2).

Currently, there are nearly 25 licensed compounds with antileishmanial effects, but only a few are used in humans. Drawbacks associated with conventional treatment with antimonials and amphotericin B include high toxicity, differences in strain sensitivity, and resistance. Moreover, the expense of these drugs often precludes their use. As recently as 2004, liposomal amphotericin B, miltefosine, and paromomycin were identified by WHO/TDR as the three most promising drugs in the market. These drugs are not new: amphotericin B has been used extensively for decades as a second-line drug for treatment of leishmaniasis (in addition to its antifungal activity), miltefosine was developed long ago as an anticancer agent, and paromomycin is more than 50 years old. To date, these three agents, together with antimonials and nonliposomal amphotericin B, are the reference chemotherapeutic agents for the leishmaniases. Oral miltefosine has been shown to be as efficacious against leishmaniasis as the standard amphotericin B treatment in India; however, important side effects, such as teratogenicity, are associated with this drug (28). With this limitation, the need for safer, inexpensive, and widely available treatments continues to be one of the top research priorities for disease control (5).

In contrast to other possible strategies (“orphan drugs,” combinatorial chemistry, or rational design), we seek new indications for existing drugs, which can be a very fruitful route for drug discovery and development (6). Pyrazinamide (PZA), an essential constituent of short-course tuberculosis chemotherapy (11), was developed as an analog of nicotinamide and used in the late 20th century in the treatment of Mycobacterium tuberculosis. PZA and related analogs have also demonstrated activity against other Mycobacterium spp. (9, 21, 27). In myco-
bacteria, we found that PZA inhibits the enzyme fatty acid synthase I (FASI) (33) by competitive inhibition of a NADPH binding site (25). By analogy, it could be proposed that PZA interferes with fatty acid synthesis in trypanosomatids. Although no genes homologous to the FASI gene have been identified in the *Leishmania* genome, this parasite employs microsomal elongases in an iterative manner to synthesize fatty acids (17). This suggests that inhibition of fatty acid synthesis might be an attractive chemotherapeutic target in *Leishmania* (16, 23).

In this article, we demonstrate that PZA has antileishmanial effect in vitro on both promastigotes and amastigotes. More importantly, PZA dramatically decreases lesion development and the parasite burden in C57BL/6 mice infected with *Leishmania major*. Finally, we show that PZA increases activation of infected macrophages and dendritic cells by increasing expression of costimulatory molecules and secretion of proinflammatory cytokines and nitric oxide. These results not only show that PZA constitutes a very promising alternative therapy for leishmaniasis but also suggest that the drug causes collateral immunostimulation.

**MATERIALS AND METHODS**

Mice. C57BL/6 mice (5 to 6 weeks of age) were purchased from Taconic (Germantown, NY). Toll-like receptor 2 (TLR-2)- and TLR-4-deficient mice (Germantown, NY). Toll-like receptor 2 (TLR-2)- and TLR-4-deficient mice were kindly provided by David Russell at Cornell University. All mice were maintained in the Baker Institute Animal Care Facility under pathogen-free conditions.

**Parasite and cell culture.** *L. major* clone V1 (MHOM/UL/89/Friedlin) promastigotes were grown at 26°C in medium 199 supplemented with 20% heat-inactivated fetal calf serum (FCS) (Gemini, Sacramento, CA), 100 U/ml penicillin, 100 μg/ml streptomycin, 2 mM L-glutamine, 40 mM HEPES, 0.1 mM adenine (in 50 mM HEPES), and 5 mg/ml hemin (in 50% triethanolamine).

The macrophage murine cell line J774 (catalog no. TIB-67) was cultured in Dulbecco’s modified Eagle medium (DMEM) (Sigma-Aldrich, St. Louis, MO) with 10% FCS, 100 U/ml penicillin, 100 μg/ml streptomycin, and 2 mM glutamine (Sigma-Aldrich) at 37°C under a 5% CO2 atmosphere. Culture medium was changed twice per week. Subcultures were performed when monolayers covered 90% of the bottom of culture flasks. For experiments involving macrophages and dendritic cells, bone marrow was obtained from C57BL/6 mouse femurs and grown for 6 to 8 days in RPMI 1640 supplemented as described above in the presence of 10% L929 conditioned medium (to generate macrophages [13]) or 20 ng/ml granulocyte-macrophage colony-stimulating factor (CSF) (to generate dendritic cells [31]).

**Drugs.** PZA was provided by Cynamon and Welch and dissolved in dimethyl sulfoxide with subsequent dilutions in water (in vivo assays) or culture medium (in vitro assays).

**Promastigote and amastigote drug treatment.** Mid-log-phase (day 3 of culture) *L. major* promastigotes were employed. Parasite concentration was adjusted to 10⁶ promastigotes/ml, and parasites were seeded into 96-well plates in a volume of 100 μl (final concentration, 10⁵ promastigotes/well). PZA was tested in triplicate in a concentration gradient from 1,000 to 0.5 μg/ml and added to the wells containing the parasite in a volume of 100 μl. A negative control was included with three wells containing only parasites and medium. The positive control consisted of amphotericin B (0.1 and 1 μg/ml; Sigma), as previously tested by us (19). After 48 h of incubation at 26°C, 10 μl of each well was diluted in 90 μl of the vital colorant (trypan blue in phosphate-buffered saline), and the parasites were quantified in a Neubauer chamber. Data were normalized as percentages of survival compared to data for untreated controls. The 50% lethal dose was extrapolated from the graph as the concentration of the products that inhibited the parasitic growth at 50% of the values of the negative control.

Amastigotes were generated by infecting either J774 murine macrophages or bone marrow-derived macrophages or dendritic cells from C57BL/6 mice. Infections were carried out after cells were seeded in eight-well Labtek chambers (Thermo Physic Scientific, Rochester, NY) at a concentration of 5 × 10⁶ cells/well. To avoid multiplication, cells were incubated with mitomycin C at a concentration of 0.8 μg/ml for 16 h. Infective-stage promastigotes of *L. major* were isolated from stationary cultures (4 to 5 days old) by Ficoll enrichment (26), added to macrophage cultures (five promastigotes to one cell), and kept overnight at 37°C in the presence of 5% CO2 and DMEM with 10% FCS. Sixteen hours later, noninternalized promastigotes were washed and replaced by culture medium containing the drug. Forty-eight hours later, wells were detached from the slides, stained with Diff-Quick (Dade Behring, Newark, DE), and counted under a light microscope. The parasite burden was determined by observation by light microscopy (magnification, ×1,000) as the number of amastigotes per 100 cells. The percentage of survival of amastigotes and concentration at which 50% of parasites were inhibited (MIC₅₀) were calculated as described above.

**J774 cell viability following incubation with PZA was also determined.** Cells were seeded on eight-well Labtek chambers as above and incubated with different concentrations of PZA (up to 1 mg/ml) for 48 h, stained with Diff-Quick, and counted under a light microscope. The percentage of viable cells was determined after quantifying the number of cells present per field (in 25 fields, ca. 500 cells). The percentage of survival and MIC₅₀ were calculated as above.

**In vivo infection and treatment studies.** Mice (*n* = 6) were anesthetized with isoflurane (Abbott Laboratories, Chicago, IL) and inhaled intradermally in both ears with 5 × 10⁴ *L. major* promastigotes in a volume of 10 μl using a 27-gauge needle. PZA was diluted in water and administered by oral gavage in a 0.2-ml volume. A control group was treated with water containing dimethyl sulfoxide (3.8%).

The timetable for the experiment was as follows: day 0, infection; days 1 to 5, 8 to 12, 15 to 19, and 22 to 26, drug administration; and day 70, sacrifice. Lesion size was monitored one to two times per week by measuring the lesion diameter with a vernier caliper. Mice were euthanized by CO2 inhalation.

**Parasite quantitation.** Parasite loads in the ears were determined as described previously (4). Briefly, the ventral and dorsal sheets of the infected ears were separated and deposited in RPMI containing 100 U/ml penicillin, 100 μg/ml streptomycin, and Liberase CI enzyme blend (0.5 mg/ml; Boehringer Mannheim). Ears were incubated for 60 min at 37°C. The sheets were dissociated using a handheld tissue homogenizer. The homogenates were filtered using a 70-μm cell strainer (BD Falcon, San Jose, CA) and serially diluted in 96-well flat-bottom microwell plate containing biphasic medium prepared using 50 μl NNN medium (25 g Bacto beef, 10 g neopeptone, 10 g Bacto agar, 2.5 g NaCl) containing 20% defibrinated rabbit blood overlaid with 100 μl medium 199. The number of viable parasites in each ear was estimated from the highest dilution at which promastigotes could be grown out after 7 days of incubation at 26°C.

**Analysis of cell activation following treatment.** J774 cells or bone marrow-derived cells were seeded on 24-well plates at a concentration of 5 × 10⁴ cells/ml. Twenty-four hours later, PZA was added to the wells at different concentrations in the absence or presence of PZA major (1:5 parasite ratio). In some experiments, J774 cells were treated with amphotericin B (0.1, 0.5, or 1 μg/ml [Sigma]). Two control groups were also included in this experiment: a positive control for activation, consisting of a group of uninfected cells treated with 100 ng/ml lipopolysaccharide (LPS) and 100 μl/ml interferon-γ (IFN-γ), and a negative control for activation, consisting of uninfected, untreated cells. Cell cultures were maintained overnight and then treated for an additional 6 h with brefeldin A (10 μg/ml), harvested by scraping, and fixed in 2% paraformaldehyde. In some experiments, flow cytometry was employed. Prior to staining of cells with fluorescent antibodies, they were incubated with an anti-Fcy III/II receptor and 10% normal mouse serum in phosphate-buffered saline containing 0.1% bovine serum albumin–0.01% NaN₃. Cells were permeabilized with saponin and stained for the expression of surface markers CD80 (clone 1D9) and inducible nitric oxide synthase (iNOS) (clone M5/114.15.2) and for the cytokines interleukin 12p40/p70 (IL-12p40/p70) (clone C17.8), IL-10 (clone JES5-16E3), and tumor necrosis factor alpha (TNF-α) (clone 1D9) and inducible nitric oxide synthase (iNOS) (clone NOS-IN). Incubations were carried out for 30 min on ice. All antibodies were purchased from BD Biosciences or eBioscience. For each sample, at least 50,000 cells were analyzed. The data were collected and analyzed using the CELLQuest software program and a FACScalibur flow cytometer (Becton Dickinson, San Jose, CA). In other experiments, supernatants from untreated cells were collected and assayed by enzyme-linked immunosorbent assay (ELISA) for the secretion of IL-12, IL-10, or TNF-α using commercial kits (Peprotech). The levels of nitric oxide in treated and untreated cultures was also measured in supernatants using the Griess reagent (Invitrogen).

**Statistical analysis.** Statistical analysis of the in vivo data used a one-way analysis of variance followed by Bonferroni’s post hoc test using the GraphPad Prism software program (San Diego, CA) (*n* = 7). Results were considered
significant when P values were ≤0.05. MIC₅₀s were interpolated from curves generated by nonlinear regression analysis using GraphPad Prism.

**RESULTS**

**PZA has antileishmanial effect in vitro.** The effect of PZA was first determined with promastigotes of *L. major* after 48 h. *L. major* promastigotes treated with PZA experienced a decrease in cell proliferation. The MIC₅₀ was established as 16.2 μg/ml (16.1 μM) (Table 1). Intracellular amastigotes appeared to be slightly more sensitive to the effect of PZA than the extracellular forms (MIC₅₀ = 10.2 μg/ml or 8.2 μM). Incubation of *L. major* with 1 μg/ml amphotericin B (as a positive control) caused 100% mortality of promastigotes and 90% of amastigotes.

To determine the potential cytotoxicity of PZA toward the mammalian cell line, coinoculation of PZA with uninfected J774 cells was also carried out for 48 h. Results showed that the drug was cytopathic at 200 μg/ml, resulting in mortality of 25% of the cell monolayer. The MIC₅₀ for mammalian cells was established as 425.6 μg/ml. The control drug amphotericin B caused only 5% mortality of the cell monolayer at the concentration tested.

To distinguish between leishmaniosstatic and leishmanicidal effects, a time course survival curve was generated for J774 cultures infected with *L. major*. Figure 1 shows parasite loads in macrophages at 24, 48, and 120 h posttreatment with 100 μM PZA. Interestingly, about 90% of *L. major* parasites were efficiently eliminated from the cells after 120 h, indicating that PZA is a leishmanicidal drug.

**PZA significantly reduces clinical disease and parasite burdens in infected mice.** To assess the efficacy of PZA in vivo, C57BL/6 mice were intradermally infected with 5 × 10⁵ *L. major* parasites in each ear (n = 6 mice; 12 ears) and treated orally with PZA at several concentrations (900 to 150 mg/kg of body weight). As shown in Fig. 2A, the oral administration of PZA produced a significant (P = 0.0001) reduction of the average lesion size in all treated groups compared with results for untreated mice at all time points. However, there were no statistical differences among the three experimental groups treated with different doses of PZA. Parasite burdens in the ears were determined at 5 weeks postinfection in the experimental groups. Figure 2B shows that treatment dramatically decreased the parasite burden in infected ears at week 6 (100-fold; P = 0.008) compared to results for the control. As before, no statistical differences were detected among the PZA-treated groups. Parasite burdens were also comparable among all experimental groups when determined after healing, at 12 weeks postinfection. Finally, PZA treatment was not toxic and did not affect the growth of the experimental animals since no significant differences in body weight were found at week seven postinfection (Fig. 2C).

**PZA increases J774 cell activation as well as release of proinflammatory cytokines and nitric oxide.** Because of the striking in vivo effect of PZA, we studied the effect of treatment on macrophages. First, we looked at activation markers and cytokine secretion by J774 cells (a murine macrophage cell line) following drug treatment (10 and 100 μM) in the presence or absence of *L. major* infection. As a positive control, cells were exposed to a mixture of IFN-γ and LPS. Figure 3 shows that drug treatment increased expression of the costimulatory molecules CD80, CD86, and MHC class II, suggesting that treatment alone increases the ability of the macrophage to present antigen. *L. major*-infected J774 cells downregulated the expression of costimulatory molecules and class II MHC molecules compared to that for infected, untreated controls, a phenomenon typically associated with *L. major* infection. Interestingly, treatment of *L. major*-infected cells with PZA rescued the ability of the cell line to upregulate all surface markers studied at the same level as that for cells treated with the drug alone.

We also determined the ability of J774 cells to produce cytokines in response to infection and/or treatment. Cells were infected, treated, or activated as described above. The amounts of the proinflammatory cytokines IL-12 and TNF-α (implicated in Th1 response and parasite killing) and the repressive

**TABLE 1. Effect of PZA on *L. major* and J774 cell survival**

<table>
<thead>
<tr>
<th>Group assessed</th>
<th>% Survival at PZA concn (μg/ml) of:</th>
<th>MIC₅₀ of PZA (μg/ml and μM)</th>
<th>% Survival with amphotericin B treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1</td>
<td>12.5</td>
<td>50</td>
</tr>
<tr>
<td>Promastigotes</td>
<td>88 ± 18</td>
<td>50 ± 18</td>
<td>38 ± 14</td>
</tr>
<tr>
<td>Amastigotes</td>
<td>89 ± 12</td>
<td>47 ± 12</td>
<td>23 ± 10</td>
</tr>
<tr>
<td>Uninfected J774 cells</td>
<td>100 ± 11</td>
<td>100 ± 10</td>
<td>100 ± 12</td>
</tr>
</tbody>
</table>

* Amphotericin B (1 μg/ml) was included in the experiment as a control. After 48 h, parasite and cell survival was determined. Data are expressed as percentages of survival compared to results for the untreated control (100% survival). Values are means ± standard deviations of at least three independent determinations.

b Value was not determined.
cytokine IL-10 in the culture supernatants were determined by ELISA. The level of nitric oxide in the same supernatants was also determined. PZA treatment alone increased cytokine production, especially that of IL-12 and TNF-α, and NO release. Cytokine production was also increased in the wells treated with PZA and infected with *L. major* compared with that for infection with *L. major* alone, suggesting again that the immune response is enhanced by PZA in infected cells (Table 2). Treatment also slightly increased IL-10 production, although the response of the cell line to the drug was dominated by the release of proinflammatory factors. Finally, to test the specificity of the immune activation by PZA, we determined the effect that amphotericin B treatment had on activation and cytokine expression by J774 cells. Treatment of J774 cells with the drug did not result in activation at the doses tested (Fig. 4).

Bone marrow-derived macrophages and dendritic cells from C57BL/6 mice also release proinflammatory factors in response to PZA. The background of J774 cells is the susceptible BALB/c mouse strain. Because BALB/c susceptibility to *L. major* is mediated by its inability to initiate Th1 responses, we...
tested the effect of the drugs on primary cells isolated from the resistant mouse strain C57BL/6.

We obtained bone marrow cells and grew them in the presence of the cytokines macrophage CSF or granulocyte-macrophage CSF to induce differentiation of macrophages or dendritic cells, respectively. In this experiment, we included the study of the immune response of dendritic cells because they are essential for the initiation the immune response against \textit{L. major}. Table 3 shows that, as before, parasite infection inhibits the initiation of inflammatory responses by macrophages, as evidenced by their inability to produce cytokines or release nitric oxide. Again, this effect was rescued by the treatment of macrophages with PZA.

Dendritic cells infected with \textit{L. major} were able to release IL-12, TNF-\(\alpha\), and nitric oxide following infection. However, this effect was greatly enhanced (10-fold) if PZA was added to the infected cells. Both dendritic cells and macrophages also increased the expression of costimulatory molecules (not shown). Together, these results suggest that PZA has immunostimulatory properties that may contribute to parasite killing beyond the leishmanicidal effect of the compounds.

The stimulatory effect of PZA is independent of TLR engagement. To confirm that the immunostimulatory effect of PZA is directly caused by the drug and is not due to contaminants (e.g., endotoxin) or other ligands that could cause cell activation via TLRs, we studied the effect of drug treatment on bone marrow-derived dendritic cells isolated from TLR-2/TLR-4 double knock-out mice. In this experiment, we treated the dendritic cells as described above in the presence or absence of \textit{L. major} infection. Again, as a positive control, cells were primed with IFN-\(\gamma\) and LPS. Figure 5A shows that drug treatment increased expression of activation markers and MHC molecules in a dose-dependent manner in dendritic cells lacking TLR-2 and -4, suggesting that PZA treatment and not a contaminant was responsible for cell activation.

\begin{table}[h]
\centering
\begin{tabular}{|c|c|c|c|c|c|}
\hline
Cytokine or substance & Drug concn \((\mu M)\) & None & \textit{L. major} & PZA & \textit{L. major}, PZA \& LPS, IFN-\(\gamma\) \\
\hline
IL-12 & 0 & 34 \pm 12 & 44 \pm 24 & 178 \pm 38* & 253 \pm 68* & 678 \pm 125* \\
& 10 & & & 359 \pm 65* & 299 \pm 55* & \\
& 100 & & & & & \\
TNF-\(\alpha\) & 0 & 135 \pm 21 & 105 \pm 19 & 245 \pm 159* & 259 \pm 163* & 1,256 \pm 132* \\
& 10 & & & 377 \pm 105* & 489 \pm 154* & \\
& 100 & & & & & \\
IL-10 & 0 & 14 \pm 39 & 54 \pm 8 & 112 \pm 45* & 109 \pm 39* & 236 \pm 22* \\
& 10 & & & 145 \pm 56* & 112 \pm 44* & \\
& 100 & & & & & \\
Nitric oxide & 0 & 37 \pm 11 & 24 \pm 18 & 254 \pm 44* & 289 \pm 55* & 921 \pm 223* \\
& 10 & & & 476 \pm 52* & 498 \pm 74* & \\
& 100 & & & & & \\
\hline
\end{tabular}
\caption{IL-12, TNF-\(\alpha\), IL-10, and nitric oxide production determined by ELISA (cytokines) or Griess test (nitric oxide) in J774 cells infected with \textit{L. major} and treated with 10 or 100 \(\mu M\) PZA
\footnote{A group of uninfected cells was treated with 100 ng/ml LPS and 10 IU IFN-\(\gamma\) as a positive control of activation. Cytokine levels \((\pm\) standard deviations) in \textit{L. major}-infected macrophages and in untreated cells were also determined. Data were obtained from three independent experiments. *, statistically significant when compared with result for untreated, \textit{L. major}-infected control group \((P < 0.05)\).}}
\end{table}

\textbf{FIG. 4.} PZA increases proinflammatory cytokine production in J774 cells. IL-10, IL-12, TNF-\(\alpha\), and nitric oxide production were determined by ELISA (cytokines) or the Griess test (nitric oxide) in J774 cells infected or not with \textit{L. major} and treated with 0.1 or 1 \(\mu M\) amphotericin B. Uninfected cells were treated with 10 U IFN-\(\gamma\) and 100 ng/ml LPS as a positive control of activation. Uninfected cell cytokine levels are shown. Data are expressed as means \pm standard deviations for three determinations. *, statistically significant; \(P = 0.0001\) compared with results for the group infected with \textit{L. major} that did not receive the drug.
significant when compared with results for untreated, L. major
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major
employed at 100
the effect of the PZA at 48 h postinfection (10-fold increase in
with a murine model of tuberculosis. Although intracellular
are comparable to what was obtained by Klemens et al. (15)
H9262
ployastigotes and 100.1
indicating that the standard antituberculous treatment regi-
now that it has been established that PZA is effective, and not
toxic, at the maximum dose proposed for humans, these initial
studies will be followed by others to determine the ideal dose/
toxications tested (900, 450, and 150 mg/ml). PZA treatment also
significantly decreased the parasite burden in the infection site
unraveled. We have previously reported that PZA inhibits the
enzyme FASI (33) by competitive inhibition of a NADPH
binding site (25). L. major lacks FASI but possesses microso-
mal elongases (17) that effect many of the same chemical
transformations as FAS. Our in vitro, and especially our in
vivo, results are consistent with our hypothesis that this path-
way is indeed an optimal target for the development of anti-
leishmanial drugs.
It has been proposed, however, that Leishmania parasites
can survive with greater altered lipid profiles than trypano-
somes and even acquire lipids from the hosting macrophage
(23). Our results demonstrate that long-term treatment of
leishmanial cultures with PZA resulted in almost complete
elimination of parasites from macrophages. If fatty acid syn-
thesis is nonessential in Leishmania or these organisms are
highly resistant to lipid depletion, it is possible that parasite
killing is not mediated exclusively by the direct effect of the
drug on parasite survival and replication. Thus, the antileish-
manial effect of PZA may be caused (or enhanced) by che-
motherapeutic interaction with the macrophage. Our data show
that J774 cells, as well as primary cells from C57BL/6 mice,
upregulate activation markers and release cytokines following
treatment with PZA, demonstrating that the drug enhances the
immune response to L. major infection. This immunoenhanc-
ing effect was not replicated in cells treated with amphotericin
B, despite publications reporting the immunostimulatory effect
of the drug (8), or in cells deficient in TRL-2 and -4 receptors,
confirming that immunostimulation is a PZA-specific event.
This phenomenon would be especially desirable in situations
where patients are immunocompromised. Because Leishmania/
HIV coinfections have been extensively documented (1), the
development of drugs that boost the immune system of the
host may be extremely useful.
The data presented here together provide the grounds for
further testing of PZA (and PZA analogs) as an antileishmanial
drug, for the determination of the immune status of the L.
major-infected mice following PZA treatment, and for exten-
sion of these screens to other models of Leishmania infection.
This pioneering alternative may work as a novel chemothera-
papeutic approach to treating leishmaniasis, in particular visceral
leishmaniasis, the most severe leishmanial disease.

DISCUSSION
The emergence of the leishmaniasis and the lack of afford-
able therapy have necessitated the development of novel anti-
leishmanial therapies. In this report, it has been shown that the
clinical antituberculous drug PZA has activity against L. major
both in vivo and in vitro. PZA is a drug that has been employed
extensively, first having been used in the treatment of pulmo-
nary tuberculosis in humans in 1949 (30). The use of a licensed,
well-known drug for indications other than the treatment of
tuberculosis could eliminate hurdles associated with the devel-
oment of new antileishmanial antigens and provide therapeu-
tic alternatives for a disease for which chemotherapy is subop-
timal. Moreover, PZA is an orally administered drug, therefore
obviating the need for parenteral injections.
Our data show that PZA is efficient at controlling the growth
of L. major in vitro. The MIC<sub>50</sub> is estimated to be 10 μM for
promastigotes and 100.1 μM for amastigotes. These MIC<sub>50</sub>s
are comparable to what was obtained by Klemens et al. (15)
with a murine model of tuberculosis. Although intracellular
amastigotes appeared to be less sensitive than promastigotes to
the effect of the PZA at 48 h postinfection (10-fold increase in
MIC<sub>50</sub>), an extended kinetic analysis revealed that PZA, em-
ployed at 100 μM, eliminated 90% of the parasites in cultured
cells after 120 h of culture. This concentration is equivalent to
what was found by Zhu et al. (32) in pharmacokinetic studies
of PZA-treated children (serum concentration was 41 μg/ml),
indicating that the standard antituberculous treatment regi-
men will be appropriate for the control of L. major infections.
This is clearly supported by our striking in vivo data, which
demonstrate that PZA treatment significantly decreased lesion
development in mice infected with L. major at all concentra-
tions tested (900, 450, and 150 mg/ml). PZA treatment also
significantly decreased the parasite burden in the infection site
without compromising the overall health of the infected mice.
Now that it has been established that PZA is effective, and not
toxic, at the maximum dose proposed for humans, these initial
studies will be followed by others to determine the ideal dose/
regimen in models of cutaneous and visceral disease.
The mechanism of action of PZA is just beginning to be
unraveled. We have previously reported that PZA inhibits the

<table>
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<tr>
<th>Cell group and cytokine or substance</th>
<th>Unstimulated</th>
<th>L. major</th>
<th>PZA (100 μM)</th>
<th>L. major, PZA (100 μM)</th>
<th>LPS, IFN-γ</th>
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<tr>
<td>Macrophages</td>
<td></td>
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<tr>
<td>IL-12</td>
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<td>34 ± 6</td>
<td>867 ± 546*</td>
<td>921 ± 445*</td>
<td>3,678 ± 456*</td>
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<td>IL-10</td>
<td>15 ± 13</td>
<td>30 ± 22</td>
<td>155 ± 21*</td>
<td>199 ± 120*</td>
<td>321 ± 156*</td>
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<td>TNF-α</td>
<td>35 ± 22</td>
<td>104 ± 89</td>
<td>758 ± 246*</td>
<td>921 ± 345*</td>
<td>2,678 ± 625*</td>
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<td>Nitric oxide</td>
<td>30 ± 3</td>
<td>12 ± 21</td>
<td>543 ± 221*</td>
<td>699 ± 112*</td>
<td>1,240 ± 516</td>
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<tr>
<td>Dendritic cells</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IL-12</td>
<td>46 ± 31</td>
<td>114 ± 26</td>
<td>956 ± 145*</td>
<td>1,035 ± 785*</td>
<td>3,365 ± 789*</td>
</tr>
<tr>
<td>IL-10</td>
<td>35 ± 33</td>
<td>160 ± 52</td>
<td>185 ± 63*</td>
<td>203 ± 60*</td>
<td>621 ± 102*</td>
</tr>
<tr>
<td>TNF-α</td>
<td>102 ± 45</td>
<td>637 ± 67</td>
<td>1,654 ± 546*</td>
<td>1,856 ± 125*</td>
<td>3,456 ± 768*</td>
</tr>
<tr>
<td>Nitric oxide</td>
<td>156 ± 39</td>
<td>243 ± 71</td>
<td>545 ± 221*</td>
<td>699 ± 212*</td>
<td>806 ± 506*</td>
</tr>
</tbody>
</table>

* A group of uninfected cells was treated with 100 ng/ml LPS and 10 IU IFN-γ as a positive control of activation. Cytokine levels (± standard deviations) in L. major-infected cells and in untreated cells were also determined. Data are expressed as pg/ml and were obtained in three independent experiments. *, statistically significant when compared with results for untreated, L. major-infected control group (P < 0.05).
ACKNOWLEDGMENT

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REFERENCES


