Lower Nitric Oxide Susceptibility of Trivalent Antimony-Resistant Amastigotes of Leishmania infantum

P. Holzmuller, D. Sereno, and J.-L. Lemesre*

UR 008 Pathogénie des Trypanosomatidés, Institut de Recherche pour le Développement, B.P. 64501, 911 Avenue Agropolis, 34394 Montpellier Cédex 5, France.

Received 21 March 2005/Returned for modification 25 April 2005/Accepted 30 June 2005

We previously documented the induction of Leishmania amastigote apoptosis by trivalent antimony (SbIII) and nitric oxide (NO). We demonstrate here that SbIII-resistant amastigotes were resistant to NO toxicity when delivered extracellularly by NO donors or intracellularly via macrophage activation. Shared biochemical targets for SbIII and NO resistance in Leishmania are discussed.

Leishmania infantum or Leishmania chagasi is responsible for canine and human visceral leishmaniasis in both the Old and the New Worlds. Leishmania parasites develop as flagellated promastigotes in the insect vector and reside as intracellular nonflagellated amastigotes in the mammalian host, which are responsible for the clinical disease manifestations. Basic treatment of leishmaniasis consists of the administration of pentavalent antimony (SbV) in the form of sodium stibogluconate or meglumine antimoniate. The mode of action of SbV implicates its reduction by the host cell to the trivalent form (SbIII) (8, 9, 21, 23). Besides this, successful chemotherapy in murine and canine models has been correlated with the efficiency of natural immunity (5, 26), and synergism between SbV and immunostimulant cytokines has been proven to be pertinent in the treatment of leishmaniasis (14, 18). We recently demonstrated that both nitric oxide (NO) and SbIII lead to amastigote cell death with some characteristics of apoptosis (11, 22). Moreover, the activity of SbIII may be directly linked to the induction of reactive oxygen intermediates such as NO (24). In order to clarify more precisely the potency of NO and SbIII, we investigated the cross-resistance of SbIII-resistant parasites to NO.

SbIII-resistant amastigotes previously described (20) were used in all experiments. The susceptibility to NO of wild-type (WT) amastigotes and amastigotes resistant to 120 μg/ml potassium antimonyl tartrate (LiSbIIIR120) was ascertained using either acidified sodium nitrite (NaNO₂) or the NO donors SNAP (S-nitroso-N-acetylpenicillamine) and DETA-NONOate ((Z)-1-[2-(2-aminoethyl)-N-(2-ammonioethyl)aminodiazlen-1-jum]-1,2-diolate). For NaNO₂, 10⁷ amastigotes per ml were incubated for 2 h at 37 °C in the dark in 0.01 M phosphate-buffered saline (PBS), pH 4.5, in the presence of 0 to 10 mM NaNO₂. After a wash in PBS, parasites were seeded in 96-well microplates at 2 x 10⁵/well in 100 μl of medium for axenic amastigotes (MAA) (13). To estimate parasite survival after NO treatment, amastigotes were cultured in the presence of 2.5 μCi of [³H]thymidine for 24 h and then filtered through GF/C filters (Whatman International) for radioactivity determination. The relative inhibition of [³H]thymidine incorporation by 50% (IC₅₀) was calculated. The NO donors SNAP and DETA-NONOate were added directly in the culture medium at concentrations ranging from 0 to 500 μM, immediately after inoculation of 2 x 10⁵ amastigotes/100 μl of MAA in 96-well microplates. After 72 h of incubation, the effective concentrations of NO donors inhibiting WT and LiSbIIIR120 amastigote growth by 50% (IC₅₀) were estimated by MTT test (21). IC₅₀ for WT and LiSbIIIR120 amastigotes are indicated in Table 1. LiSbIIIR120 amastigotes were 2.5-, 3.2-, and 3.6-fold more resistant than the WT to NO delivered by NaNO₂, SNAP, and DETA-NONOate. NO-mediated Leishmania amastigote killing was also determined using impermeable DNA intercalant YOPRO-1 staining analyzed by a FACScan flow cytometer (Becton Dickinson, Ivy, France) (11, 22). Green cell fluorescence could selectively differentiate viable (Fig. 1a and e) and apoptotic (Fig. 1b and f) amastigotes by using the combined analysis of their forward scatter (FSC-H) and FL1-H (525 nm band pass filter) patterns. Both WT and LiSbIIIR120 amastigotes incubated in MAA alone displayed a homogeneous population of living cells (data not shown). When amastigotes were treated with 500 μM NaNO₂, a new cell population with a high FL1 fluorescence corresponding to apoptotic cells was detected for the WT (Fig. 1c and g), whereas more than 80% living cells were recorded for LiSbIIIR120 amastigotes (Fig. 1d and h).

### TABLE 1. Cytotoxic effects of NO donors on WT amastigotes of L. infantum and amastigotes resistant to 120 μg/ml potassium antimonyl tartrate LiSbIIIR120

<table>
<thead>
<tr>
<th>NO donor</th>
<th>WT amastigotes</th>
<th>LiSbIIIR120 resistant amastigotes</th>
</tr>
</thead>
<tbody>
<tr>
<td>NaNO₂</td>
<td>315.9 ± 5.3</td>
<td>6,785.9 ± 1133</td>
</tr>
<tr>
<td>SNAP</td>
<td>72 ± 0.8</td>
<td>229.1 ± 15.9</td>
</tr>
<tr>
<td>DETA/NONOate</td>
<td>37.7 ± 4.25</td>
<td>134.2 ± 2.3</td>
</tr>
</tbody>
</table>

* Corresponding author. Mailing address: UR 008 Pathogénie des Trypanosomatidés, IRD (Institut de Recherche pour le Développement), B.P. 64501, 911 avenue Agropolis, 34394 Montpellier Cédex 5, France. Phone: 33 (0)4 67 41 62 20. Fax: 33 (0)4 67 41 63 30. E-mail: lemesre@mpl.ird.fr.
LiSbIIIR120 amastigotes were also able to resist NO-mediated cell death in an intracellular position. A human leukemia monocyte cell line (THP-1 cells) was differentiated in 16-well LabTek tissue culture slides (Nalge Nunc International) (25), infected with stationary-phase *L. infantum* extracellular amastigotes (parasite-macrophage ratio of 3:1 for 2 h at 37°C with 5% CO₂), and activated by lipopolysaccharide (10 ng/ml) and gamma interferon (100 U/ml). Culture supernatants were collected 48 h later for nitrate and nitrite measurement (19), and macrophages were fixed with methanol and stained with Giemsa for parasite counts or processed for apoptosis determination (terminal deoxynucleotidyltransferase-mediated dUTP-biotin nick end labeling [TUNEL] technique, in situ colorimetric apoptosis detection system; Promega, Madison, Wis.). The parasitic index (PI) was calculated as follows: PI = mean number of amastigotes per macrophage × percentage of infected macrophages × 100. A PI reduction of more than 95%, correlated with a nitrate-nitrite accumulation indicative of NO production, was observed for WT amastigotes inside activated THP-1 macrophages (Fig. 2A). When THP-1 cells were infected with LiSbIIIR120 amastigotes, no such PI reduction was noticed (Fig. 2A). The in situ TUNEL analysis revealed that apoptosis of intracellular amastigotes only occurred in WT-infected activated THP-1 macrophages (Fig. 2B and C).

Our observation that in vitro-selected, SbIII-resistant amastigotes were cross-resistant to NO raised the questions of the existence of shared biochemical targets for SbIII and NO resistance in *Leishmania* and of the occurrence of such phenomenon in areas of endemicity.

Among the mechanisms responsible for antimony resistance in vitro, implication of trypanothione and an As-thiol pump is well documented (1, 6, 17). Interestingly, thiol groups of proteins are currently considered as targets for NO (4, 16). Thus, one explanation for NO resistance in SbIII-resistant amastigotes should be an increased content of trypanothione and thiol-related compounds (3). Extrusion of nitrosothiols by an AS-thiol pump could participate in reducing the detrimental action of NO. A recent finding showed that SbIII was able to stimulate the host immune system, leading to potential synergistic effects between the chemical and natural microbicidal molecules (24). The authors suggest that antimonial drug treatment induced the production of nitrogen species by macrophages and can cause apoptosis of intracellular amastigotes. The involvement of NO in the cytotoxic effect of SbIII could explain the lower susceptibility of SbIII-resistant amastigotes to NO action and could greatly help in the understanding of mechanism underlying antimony resistance.

Increasing cases of SbV treatment failure have been described (2, 7, 10, 12, 15), suggesting that antimony-resistant parasites could be transmitted even in hosts developing an efficient immune response. These data strongly suggest that NO-SbIII cross-resistance could have a dreadful implication for the spread of chemoresistance in the field. In this view, it will be of interest to determine whether or not SbIII-resistant strains isolated from relapsed patients were NO cross resistant.
This work was supported by grants from the Institut de Recherche pour le développement (IRD Institute) and the Fondation pour la Recherche Médicale (FRM Foundation).

REFERENCES


